

Leia Summer 6/8-6/10

WEDNESDAY, 6/8/2022

- Clean and concentrate Jeffrey's PCR products
 - Labels:
 - 3 - 13.2 ng/uL tdkkan
 - 6 - 9.70 ng/uL tdkkan

THURSDAY, 6/9/2022

- Made 1 bag of LB/KAN plates
- Resuspended and made working stocks of iGEM_005 to 012 primers (in Froslass)
- While waiting around:
 - <https://tpwd.texas.gov/huntwild/wild/diseases/whitenose/>
 - <https://austinbatrefuge.org/education-materials/>
 - Merlin Tuttle, a renowned bat conserationist and scientist: <https://www.merlintuttle.org/faq/>
 - Edited some wiki stuff

FRIDAY, 6/10/2022

- Competent cells (NEB alpha)
 - OD (1 mL): 0.405
 - Aliquots of 50 mL instead of 35 mL
 - Centrifuge 6 C instead of 4, 3000 rcf instead of 3400
 - Making 500 mL of 100 mM CaCl₂ 10% glycerol buffer:
 - 5.05 g CaCl₂
 - 50 mL glycerol
 - 500 mL - 50 mL - 5 mL = 445 mL of Milli-Q H₂O
 - USED CAMERON'S 100 mM BUFFER INSTEAD, since it was iced and cold
 - When resuspending, split 150 mL into 3 50 mL Falcon tubes (so 3 pellets)
 - When resuspending the second time, combine 3 pellets into 1 and total of 20 mL buffer
 - Snap freezing
 - Microcentrifuge tubes with comp cells marked with ORANGE SHARPIE STRIPE (looks reddish orange)! Some have black thin Sharpie writing with date
- When done centrifuging, make sure to return temperature to 25 C and air it out
- Comp cells in the unlabeled box in -80 freezer, same column as iGEM box
 - Leftover comp cells in the column behind the iGEM column (under the BTX boxes)