

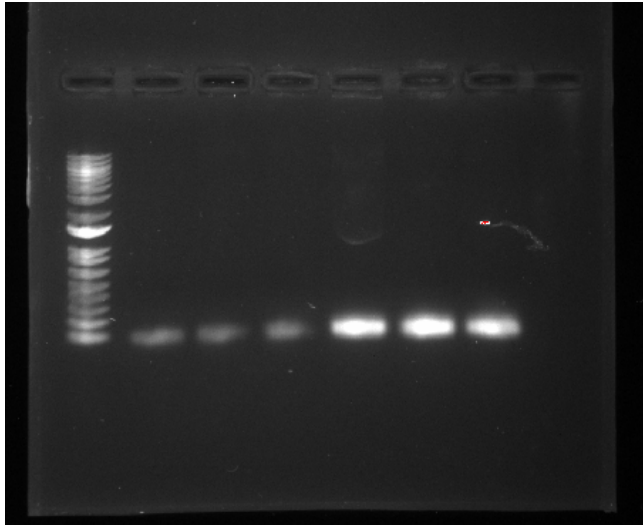
# Jeffrey Chuong 7/5 - 7-10

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TUESDAY, 7/5/2022

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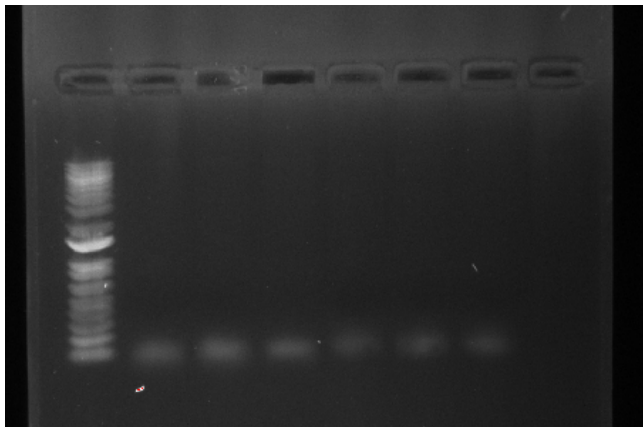
📎 7-5-22 recJ 55.8.png



*recJ* PCR: Annealing temp 55.8C for upstream and downstream

*recJ* Gel: -, Up, Up, -, Down, Down

📎 7-5-22 pbpG 57.4.png



*pbpG* PCR: Annealing temp 57.4C for upstream and downstream

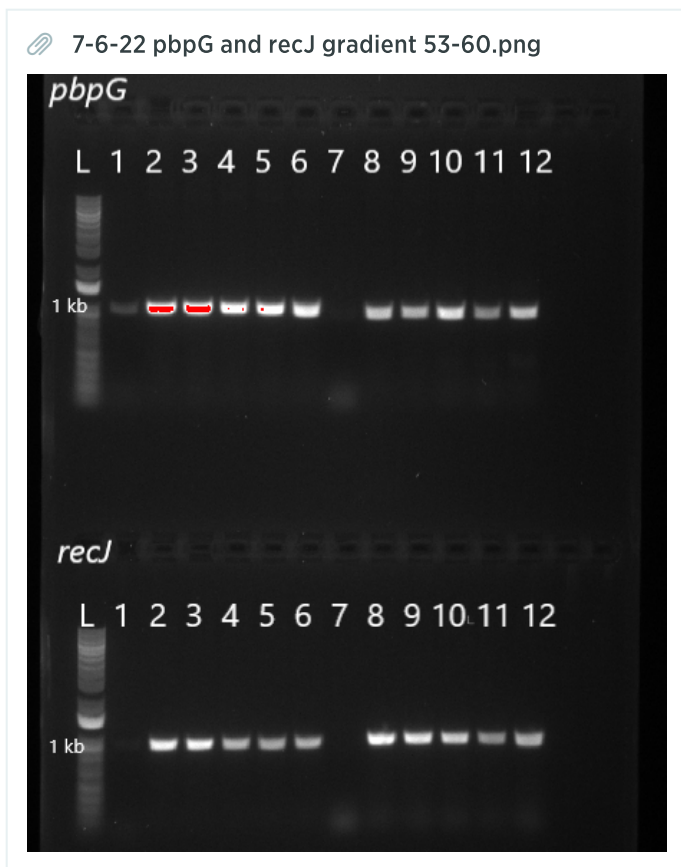
*pbpG* Gel: -, Up, Up, -, Down, Down

WEDNESDAY, 7/6/2022

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REPEAT PCR - made new primer stocks

- 53 to 60 for both annealing temperatures



Sequencing results analysis: Matt thinks the Sanger sequencing by Eton is just bad, I shouldn't trust the results - will talk to Dr. Mishler

Started GGAs for *recJ* and *pbpG* - *recJ* homologies ligated with *tdk/kan*, *pbpG* homologies ligated together

#### THURSDAY, 7/7/2022

*recJ* - made 500 ul transformations with 35 ul O/N culture ADP1-ISx and 20 ul GGA ligation of *recJ* homologies with pBTK622 *tdk/kan*

*pbpG* - made 500 ul transformations with 35 ul O/N culture iGEM22\_012 and 20 ul GGA ligation of *pbpG* homologies

Incubated O/N 30C 200 RPM

#### FRIDAY, 7/8/2022

Made new LB-KAN plates

- old LB-KAN plates were faulty or only 10 mL, could have been causing the issues for Leia?
- suggestions to defeat condensation

*recJ* plating +DNA and -DNA transformations on LB-Kan - diluted 100x in sterile saline, plated 50 ul

*pbpG* plating +DNA and -DNA transformations on LB-AZT - diluted 100x in sterile saline, plated 50 ul

#### SATURDAY, 7/9/2022

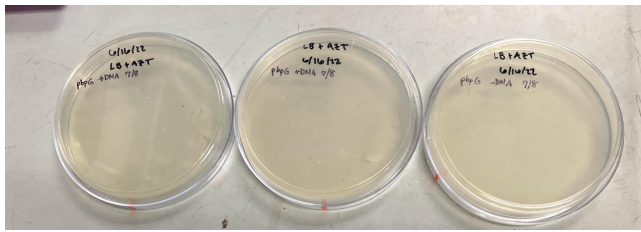
7-9-22 Transformation of ADP1-ISx with GGA ligation of *recJ* flanks with pBTK622 tdk kan.jpg



*recJ*

- Inoculated two colonies from each successful +DNA plate, may have messed up on some cultures, liquid LB-Kan cultures marked with an \* are good
- May try whole cell PCR to confirm *tdk/kan* insertion with *recJ* upstream forward and downstream reverse primers (3.7 kb)
- Make a glycerol stock tomorrow

7-9-22 Transformation of iGEM\_012 with ligated *pbpG* rescue flanks Day 1.jpg



*pbpG*

- Did not inoculate colonies due to very little growth, may try leaving it an extra day and try with the colonies that show up
- I need to check primer restriction sites on *pbpG* primers