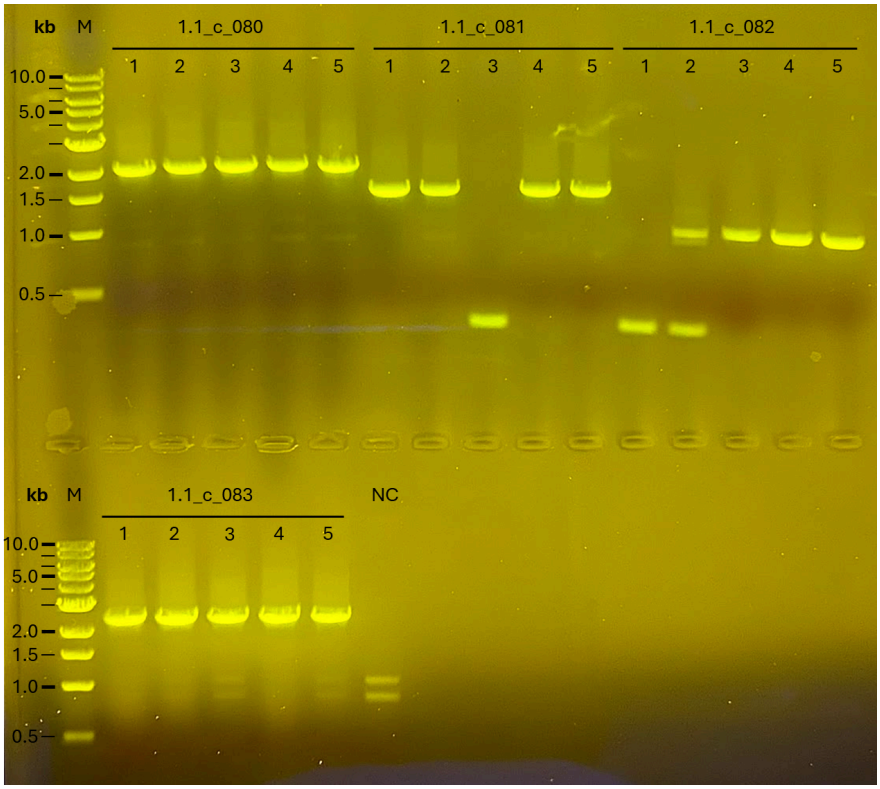
Lvl1.1_c_080; Lvl1.1_c_081; Lvl1.1_c_082; Lvl1.1_c_083 from 27.07



Lvl1.1_c_080 - 2108 bp
Lvl1.1_c_081 - 1574 bp
Lvl1.1_c_082 - 986 bp
Lvl1.1_c_083 - 2564 bp

- [x] E. coli Night Seq of Alex John Thottumkal :
  Lvl1.1_c_080 - 3 (fw + rv)
  Lvl1.1_c_081 - 1 (fw + rv)
  Lvl1.1_c_082 - 5 (fw)
  Lvl1.1_c_083 - 4 (fw + rv)
  Lvl1.3_c_086 - 4 (fw + rv) from 22.07.2026

- [x] Trafo of Golden Gate reactions from 28.7.2026 according to E. coli Transformation Ziyue Zhou
  Lvl1.1_c_084 28.7.
  Lvl1.1_c_085 28.7.

DONNERSTAG, 30.7.2026

cPCR of Golden Gate from 28.7.2026: Very few colonies were observed. Incubate for another 24 hours. Ziyue Zhou

- [x] Redo trafo of Golden Gate reactions from 28.7.2026 according to E. coli Transformation Ziyue Zhou
  Lvl1.1_c_084 28.7.
  Lvl1.1_c_085 28.7.

For level 1: Spread all the bacteria onto **one** agar plate, instead of splitting them onto two plates like the protocol says.

FREITAG, 31.7.2026

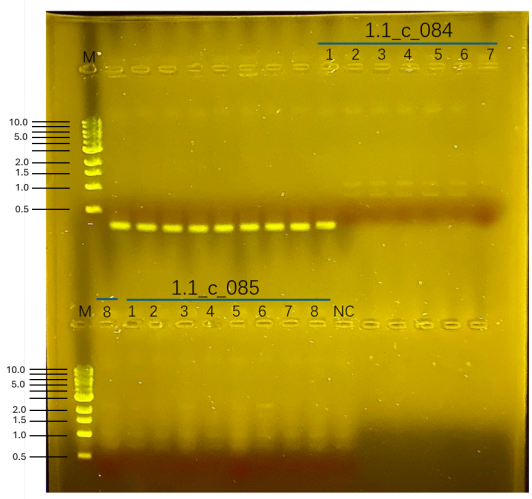
- [x] cPCR of Golden Gate from 28.7.2026 according to Taq colony PCR Ziyue Zhou
  Lvl1.1_c_084 28.7.
  Lvl1.1_c_085 28.7.

Mastermix preparation Taq cPCR. Level 1.1 (the rest of the modules, see above)

	A	B	C	D
1		50ul Reaction	**25ul Reaction**	**xMM**
2				
3	2x Taq MM	25ul	12.5	12.5
4	10uM P fw	1ul	0.5	0.5
5	10um P rv	1ul	0.5	0.5
6	Template	colony	colony	colony
7	H20	to 50ul	11.5	11.5
8				

- [x] gel electrophoresis of cPCR from 28.7.2026 Ahmed Haj Kacem Alex John Thottumkal
  Lvl1.1_c_084 28.7.
  Lvl1.1_c_085 28.7.

Lvl1.1_c_084; Lvl1.1_c_085 from 28.07



MONTAG, 3.8.2026

- [x] Redo cPCR of trafo from 29.7 and 30.7 according to Taq colony PCR Ziyue Zhou
  Lvl1.1_c_084 28.7.
  Lvl1.1_c_085 28.7.

Mastermix preparation Taq cPCR. Level 1.1 redo modules 84 and 85

	A	B	C	D
1		50ul Reaction	**25ul Reaction**	**14MM**
2				
3	2x Taq MM	25ul	12.5	175
4	10uM P fw	1ul	0.5	7
5	10um P rv	1ul	0.5	7
6	Template	colony	colony	colony
7	H20	to 50ul	11.5	161
8				

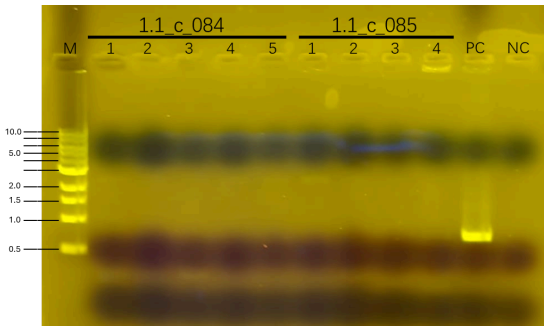
DIENSTAG, 4.8.2026

- [x] Gel electrophoresis of cPCR 03.08 Ziyue Zhou

  Lvl1.1_c_084 28.7.

  Lvl1.1_c_085 28.7.

Lvl1.1_c_084; Lvl1.1_c_085 from 28.7.



- [x] Redo Golden Gate according to Golden Gate Assembly Ziyue Zhou

  Lvl1.1_c_084 28.7.

  Lvl1.1_c_085 28.7.

Master Mix Preparation for Golden Gate Assembly. Redo modules 84 and 85

	A	B	C	D	E
1		reaction PCR amplified or ordered (15nM stock)	4x MM	2x MM	
2	each Level 0 plasmid 15nM	1	4	8	
3	10x T4 Ligase Buffer	1	4	2	
4	T4 Ligase	0.5	2	1	
5	BsaI (10 U)	0.5	2	1	
6	Backbone (1 µl of 15nM stock)	Make up to 10 µl with 15nM Backbone		6	
7	PABP			2	

- [x] Set up pre-culture for yeast growth curve Ziyue Zhou

MITTWOCH, 5.8.2026

- [x] Trafo of Golden Gate 04.08 according to E. coli Transformation Ziyue Zhou

  Lvl1.1_c_084 28.7.

  Lvl1.1_c_085 28.7.

- [x] pre-culture growth check Ziyue Zhou

DONNERSTAG, 6.8.2026

- [x] cPCR of Golden Gate 4.8 according to Taq colony PCR Alex John Thottumkal

  10 colonies were picked, blue positive control was picked from Lvl1.1_c_084 plate

  Lvl1.1_c_084 28.7.

  Lvl1.1_c_085 28.7.

Master Mix Preparation Taq cPCR. Level 1.1 redo modules 84 and 85

	A	B	C	D
1		50ul Reaction	**25ul Reaction**	26x MM
2				
3	2x Taq MM	25ul	12.5	325
4	10uM P fw	1ul	0.5	13
5	10um P rv	1ul	0.5	13
6	Template	colony	colony	colony
7	H20	to 50ul	11.5	299
8				

- [x] gel electrophoresis of Golden Gate 4.8 Alex John Thottumkal

  Lvl1.1_c_084 28.7.

  Lvl1.1_c_085 28.7.

  insert gel pic

  Lvl1.1_c_084 - 2030 bp

  Lvl1.1_c_085 - 1442 bp
- [x] yeast growth curve day 1 Ziyue Zhou

  bad results

FREITAG, 7.8.2026

- [x] yeast growth curve day 2 Ziyue Zhou

  bad results

MONTAG, 10.8.2026

- [x] set up overnight of following for glycerol stocks, but check sequencing results Ahmed Haj Kacem :

  1:1000 Spec

  3 mL Medium, 3 µL of antibiotic

  Lvl1.1_c_080 - 3 (fw + rv)

  Lvl1.1_c_081 - 1 (fw + rv)

  Lvl1.1_c_082 - 5 (fw)

  Lvl1.1_c_083 - 4 (fw + rv)
- [x] Redo E. coli Nightseq of Ahmed Haj Kacem Alex John Thottumkal :

  Lvl1.3_c_086 - 4 fw (this time only fw)

  and pick new colonies form 086 and run E. coli Night Seq with fw and rv
- [x] Re-do Golden Gate Level 1 of Ahmed Haj Kacem :

  1.1_c_084

  1.1_c_085

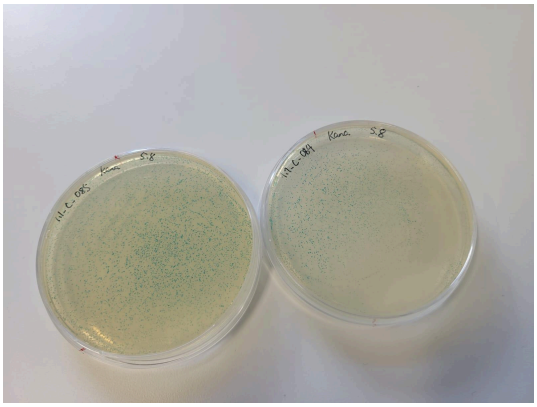
Master Mix preparation Golden Gate Assembly. Level 1.1 Redo modules 84 and 85

	A	B
1	Construct	2.2x
2	0.1_c_ 056 INT	2.2
3	0.2_a_010 L1	2.2
4	0.4_a_021 NCS rc	2.2
5	0.5_a_023 NCS rc	2.2
6	10x T4 Ligase Buffer	2.2
7	T4 Ligase	1.1
8	BsaI	1.1
9	Lvl 1 BB	6.6

DIENSTAG, 11.8.2026

- [x] yeast growth curve day 1 Ziyue Zhou Tim
- [x] in order to minimize bacterial growth on the plates (too many blue colonies on the plates from 06.08.2026).... Ahmed Haj Kacem

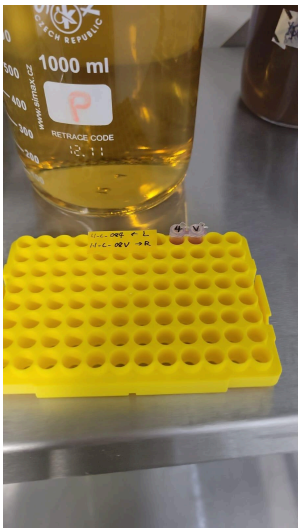
Plates_1.1_c_084_und_1.1_c_085_2026-08-06_at_09.09.29.jpeg



....plate Golden Gate reactions of 1.1_c_084 and 1.1_c_085 according to E. coli Transformation again BUT this time:

- 100 µL vorm Herunterzentrifugieren ausplattieren auf eine Platt (Schritt 8 im E. coli Transformationsprotokoll) ausplattieren
- 20 µL nachm Herunterzentrifugieren ausplattieren auf zweite platte
- Rest danach auch ausplattieren auf dritte platte

GoldenGateReactions 1.1_c_084_085_2026-08-07.jpeg



<- golden gate reaction tubes are in the freeze room (nothing left in there, redo Golden Gate Level 1 of 084 and 085)

MITTWOCH, 12.8.2026

- [x] yeast growth curve day 2 Ziyue Zhou Tim
- [x] set up overnight cultures in the afternoon. pick the (circled number) colonies from masterplate. Masterplate can be found in the 37 °C incubator Ahmed Haj Kacem

    Lvl1.1_c_080 - 3 (fw + rv)

    Lvl1.1_c_081 - 1 (fw + rv)

    Lvl1.1_c_082 - 5 (fw)

Lvl1.1_c_083 - 4 (fw + rv)

- [x] cPCR Lvl1.1_c_084 and Lvl1.1_c_085 from 11.08.2026
  - Lvl1.1_c_084 - 2030 bp
  - Lvl1.1_c_085 - 1442 bp

6 colonies were picked per construct

primers: oSN 17 & 18

Master Mix Preparation for Taq cPCR of Level 1.1 modules 84 and 85

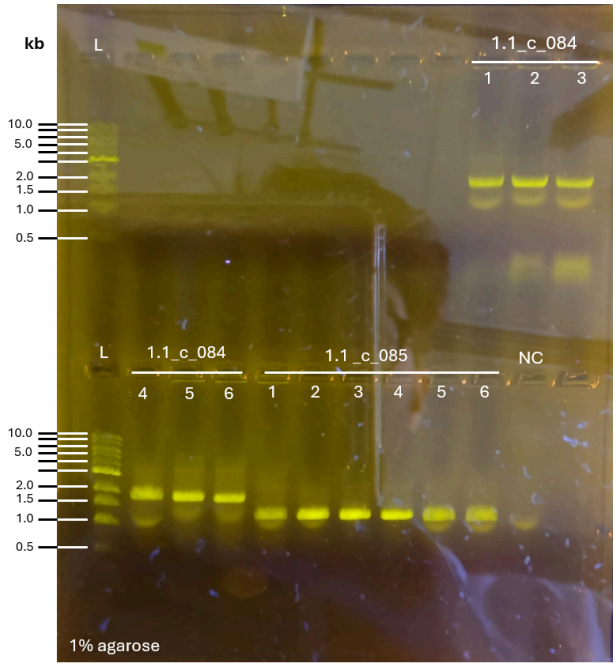
	A	B	C
1		**25ul Reaction**	13.2 x MM
2			
3	2x Taq MM	12.5	165
4	10uM P fw	0.5	6.6
5	10um P rv	0.5	6.6
6	Template	colony	colony
7	H20	11.5	151.8

PCR cycle

	A	B	C
1	STEP	TEMP	TIME
2	Initial Denaturation	95°C	30 seconds
3	25-33 Cycles	95°C 45-68°C* 68°C	15-30 seconds 15-60 seconds 2min 10s
4	Final Extension	68°C	5 minutes
5	Hold	4-10°C	

- [x] gel electrophoresis

Gel electrophoresis results 12.08.2026



The positives, that would be sent to E. coli NightSeq: **1.1_c_084_2 & 1.1_c_085_4**

DONNERSTAG, 13.8.2026

from the overnight culture from 12.8.2026:

- [x] glycerol stock of 500 µL of overnight culture with 500 µL glycerol in cryo-vial: Tim
  - Lvl1.1_c_080 - 3 (fw + rv)
  - Lvl1.1_c_081 - 1 (fw + rv)
  - Lvl1.1_c_082 - 5 (fw)
  - Lvl1.1_c_083 - 4 (fw + rv)

from the overnight culture from 12.8.2026:

- [x] Miniprep of: Tim
  - Lvl1.1_c_080 - 3 (fw + rv)
  - Lvl1.1_c_081 - 1 (fw + rv)
  - Lvl1.1_c_082 - 5 (fw)
  - Lvl1.1_c_083 - 4 (fw + rv)
- [x] Nanodrop Ahmed Haj Kacem
  - Lvl1.1_c_080 - 3 (fw + rv)
  - Lvl1.1_c_081 - 1 (fw + rv)
  - Lvl1.1_c_082 - 5 (fw)
  - Lvl1.1_c_083 - 4 (fw + rv)

- [x] Level 2 Golden Gate afternoon, thermocycler runs overnight according Golden Gate Assembly Ahmed Haj Kacem
  - Lvl.2_c_091
  - Lvl.2_c_092
  - Lvl.2_c_093
  - Lvl.2_c_094 (will not be used moving forward, wrong assembly INT + GAL + EncQt)

Master Mix Preparation Golden Gate Assembly Level 2 module 091-094. (94 wrong assembly)

	A	B	C
1	each Level 1 plasmid 15nM (3 inserts)	3	4.2 x MM
2	10x T4 Ligase Buffer	1	4.2
3	T4 Ligase	0.5	2.1
4	BbsI	0.2	0.84
5	Backbone (1 µl of 15nM stock)	5.3	22.26

BEFORE 17:00

- [x] Re-Do E. coli NightSeq of: Tim

  1.3_c_086_5 fw with forward primer oSN17
  - pick colony 5 from masterplate that has the streak 1.3_c_086_5
  - spin down E. coli NightSeq Tube (are in the cooling room) in the centrifuge
  - then pick the colony 5 1.3_c_086_5, swirl in the NightSeq tube and pipette up&down thoroughly
  - in a separate Eppi tube, pipette 20 µL of oSN17 primer (10 µM). Label it with a yellow tape and write on it " Primer oSN17"

  1.1_c_084_2 & _085_4 positives from gel 12.08.2026

FREITAG, 14.8.2026

MONTAG, 17.8.2026

1.1_c_084_2 1.1_c_085_4. Regarding 1.3_c_086_5 the sequencing result showed: NNNNN.

- [x] overnight culture of 1.1_c_084_2 1.1_c_085_4 from masterplate 12.08.2026 Ahmed Haj Kacem
- [x] E. coli trafo according to E. coli Transformation and let incubate in 37 C overnight of: Ziyue Zhou

  Lvl.2_c_091
  Lvl.2_c_092
  Lvl.2_c_093
  Lvl.2_c_094 (will not be used moving forward, wrong assembly INT + GAL + EncQt)

DIENSTAG, 18.8.2026

- [x] glycerol stock of the overnight culture from 17.08.2026: 1.1_c_084_2 1.1_c_085_4 Ahmed Haj Kacem
- [x] Miniprep of 1.1_c_084_2 1.1_c_085_4 Ahmed Haj Kacem
- [x] Nanodrop of plasmid DNA: 1.1_c_084_2 1.1_c_085_4 Ahmed Haj Kacem
- [x] overnight culture of 1.3_c_086_5 from 18.08.2026

MITTWOCH, 19.8.2026

- [x] colony PCR according to Taq colony PCR of: Ziyue Zhou

  Lvl.2_c_091
  Lvl.2_c_092
  Lvl.2_c_093

find out what primers are needed to get into the level 2 backbone!!

Master Mix Preparation Taq cPCR Level 2 modules: 91 - 93

	A	B	C
1		**25ul Reaction**	21.6 x MM
2			
3	2x Taq MM	12.5	270
4	10uM P fw	0.5	10.8
5	10um P rv	0.5	10.8
6	Template	colony	colony
7	H20	11.5	248.4

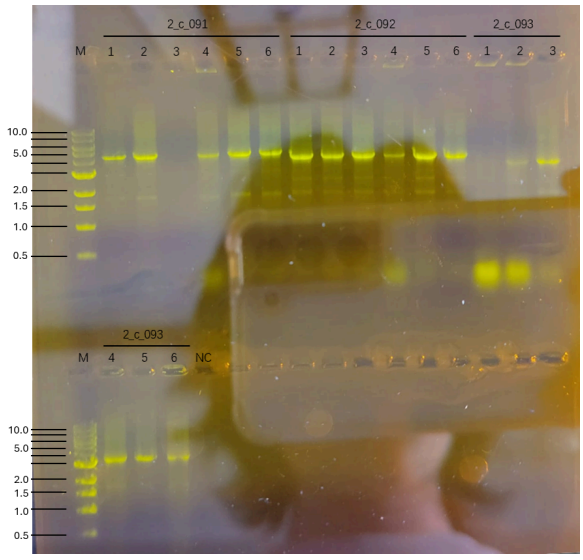
- [x] glycerol stock of 1.3_c_086_5 from 19.08.2026 Ahmed Haj Kacem
- [x] miniprep of 1.3_c_086_5 from 19.08.2026 Tim
- [x] overnight culture 1.3_c_086_02 und 1.3_c_086_04 Ahmed Haj Kacem

DONNERSTAG, 20.8.2026

- [x] gel electrophoresis of: Ziyue Zhou

  Lvl.2_c_091
  Lvl.2_c_092
  Lvl.2_c_093

Taq cPCR: Lvl2_c_091, 092 & 093



- [x] Redo cPCR with Q5 polymerase according to Q5 PCR of: Ziyue Zhou

  Lvl2_c_091_1
  Lvl2_c_092_2
  Lvl2_c_093_5

Master Mix Preparation Q5 cPCR Level 2 redo modules 91 - 93

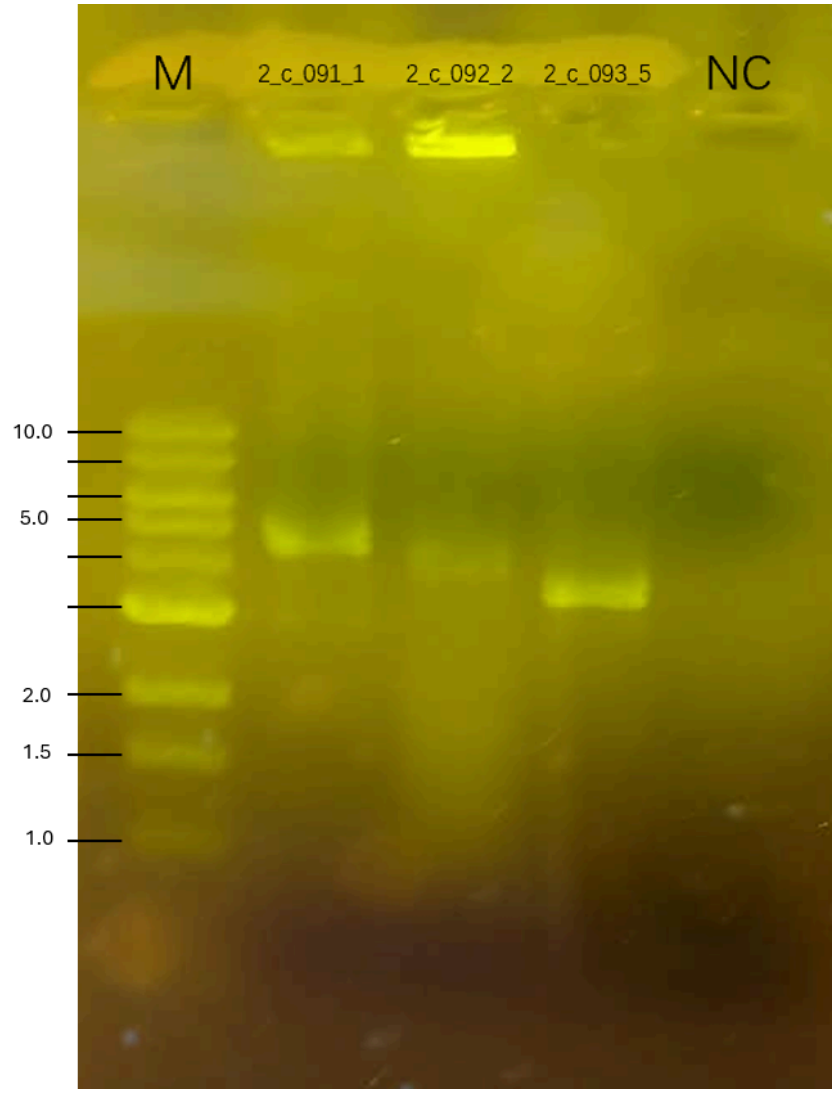
	A	B	C	D	E	F	G
1		25ul Reaction	50ul Reaction	12.5ul Reaction	4.2x 12.5	11x12.5	
2	Q5 MM	12.5	25	6.25	26.25		
3	10uM P fw	1.25	2.5	0.625	2.625		
4	10uM P rv	1.25	2.5	0.625	2.625		
5	Template (1ng)	1	1				
6	Nuclease-free H20	9	19	5	21		
7							

- [x] Miniprep 1.3_c_086_02 und 1.3_c_086_04 Ahmed Haj Kacem
- [x] Nanodrop 1.3_c_086_02 und 1.3_c_086_04 und 1.3_c_086_5 Ahmed Haj Kacem

FREITAG, 21.8.2026

- [x] Gel electrophoresis of Ziyue Zhou
  - Lvl2_c_091_1
  - Lvl2_c_092_2
  - Lvl2_c_093_5

Q5 cPCR: Lvl2_c_091_1, Lvl2_c_092_2, Lvl2_c_093_5



Prep for Sequencing Lvl 0 Send to Sequencing (Mycrosynth) Alex John Thottumkal :

12µl of plasmid eluated containing 40-100 ng/µl

3 µl of 20uM 69_level0_seq_fw

Economy run in 1.5 Eppis

in a separate Eppi tube, pipette 20 µL of oSN17 primer (10 µM). Label it with a yellow tape and write on it " Primer oSN17"

Sequencing 21.08

	A	B	C	D	E	F	G	H	I
1		seq number	construct	ng/µL	A260/A280	A260/A230	plasmid ul	plasmid amount	Water amount
2	1	98902337	1.3_c_086_2	219.4	1,86	2,39	3	54.85	9
3	2	98902338	1.3_c_086_4	335.9	1,90	2,34	2	55.9833333333	10
4	3	98902339	1.3_c_086_5	230.3	1,87	2,40	3	57.575	9
5	4								
6	5	98902329	0.3_c_060_PABP_RRM1,2	492.2	1.89	2.29	2	82.0333333333	10

MONTAG, 24.8.2026

- [x] set up overnight culture of Ziyue Zhou
  - Lvl2_c_091_1
  - Lvl2_c_092_2
  - Lvl2_c_093_5

  16:00: Incubator runs at yeast mode due to the pre-existing yeast culture inside.

DIENSTAG, 25.8.2026

- [x] Miniprep of: Ziyue Zhou
  - Lvl2_c_091_1
  - Lvl2_c_092_2
  - Lvl2_c_093_5
- [x] prep for sequencing of Send to Sequencing (Mycrosynth) Ziyue Zhou
  - Lvl2_c_091_1
  - Lvl2_c_092_2
  - Lvl2_c_093_5

Nanodrop 25.8

	A	B	C	D
1		c (ng/µL)	A260/280	A260/230
2	Lvl2_c_091_1	73.3	1.93	1.99
3	Lvl2_c_092_2	55.2	1.89	1.92
4	Lvl2_c_093_5	71.2	1.89	1.99

Sequencing result is good :)

- [x] Level 2 Golden Gate Golden Gate Assembly Ahmed Haj Kacem
  - Lvl2_c_094 (1.1_c_083 + 1.2GAL1,10 P + 1.3_c_086)
  - Lvl2_c_095 (1.1_c_084 + 1.2GAL1,10 P + 1.3_c_086)
  - Lvl2_c_096 (1.1_c_085 + 1.2GAL1,10 P + 1.3_c_086)

  Note: Lvl 1.3_c_086_2 wird benutzt

Master Mix Preparation for Golden Gate Assembly Level 2 modules: 94 - 96

	A	B	C
1	each Level 1 plasmid 15nM	3	MM x 3.2
2	10x T4 Ligase Buffer	1	3.2
3	T4 Ligase	0.5	1.6
4	BbsI	0.2	0.64
5	Backbone (1 µl of 15nM stock)	5.3	16.96

- [x] overnight culture of Lvl 1.3_c_086_2 Ahmed Haj Kacem

MITTWOCH, 26.8.2026

- [x] Glycerol stock of Lvl 1.3_c_086_2 Ahmed Haj Kacem
- [x] Transformation of E.coli with: E. coli Transformation Alex John Thottumkal
  - Lvl2_c_094 (1.1_c_083 + 1.2GAL1,10 P + 1.3_c_086)
  - Lvl2_c_095 (1.1_c_084 + 1.2GAL1,10 P + 1.3_c_086)
  - Lvl2_c_096 (1.1_c_085 + 1.2GAL1,10 P + 1.3_c_086)

DONNERSTAG, 27.8.2026

- [x] cPCR of: Taq colony PCR Alex John Thottumkal
  - Lvl2_c_094 (1.1_c_083 + 1.2GAL1,10 P + 1.3_c_086)
  - Lvl2_c_095 (1.1_c_084 + 1.2GAL1,10 P + 1.3_c_086)
  - Lvl2_c_096 (1.1_c_085 + 1.2GAL1,10 P + 1.3_c_086)
- [x] set up overnight of : Ahmed Haj Kacem
  - Lvl2_c_091_1
  - Lvl2_c_092_2
  - Lvl2_c_093_5

FREITAG, 28.8.2026

- [x] take out masterplate of incubator, put parafilm around it and store at cooling room
  - Lvl2_c_094 (1.1_c_083 + 1.2GAL1,10 P + 1.3_c_086)
  - Lvl2_c_095 (1.1_c_084 + 1.2GAL1,10 P + 1.3_c_086)
  - Lvl2_c_096 (1.1_c_085 + 1.2GAL1,10 P + 1.3_c_086)

(if positive)

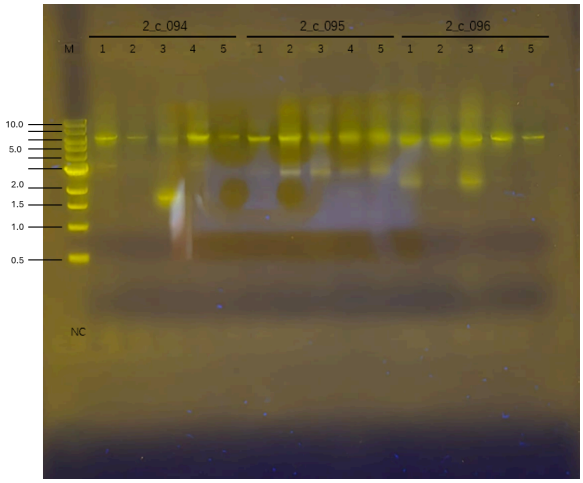
- [x] glycerol stock of: Ahmed Haj Kacem
  - Lvl2_c_091_1
  - Lvl2_c_092_2
  - Lvl2_c_093_5
- [x] Miniprep of: Tim
  - Lvl2_c_091_1
  - Lvl2_c_092_2
  - Lvl2_c_093_5
- [x] Level 2 yeast transformation of: S. cerevisiae Transformation Ahmed Haj Kacem
  - Lvl2_c_091_1
  - Lvl2_c_092_2
  - Lvl2_c_093_5

  erister transformierte Level 2er yeahhh o(*￣▽￣*)ブ

- [x] gel electrophoresis of: Ziyue Zhou
  - Lvl2_c_094 (1.1_c_083 + 1.2GAL1,10 P + 1.3_c_086)
  - Lvl2_c_095 (1.1_c_084 + 1.2GAL1,10 P + 1.3_c_086)
  - Lvl2_c_096 (1.1_c_085 + 1.2GAL1,10 P + 1.3_c_086)

  reduzierte Spannung: 75V und Gelkonz.: 0,8%

Taq cPCR: Lvl2_c_094-095-096



MONTAG, 31.8.2026

- [x] E. coli Nightseq of: Ahmed Haj Kacem
  - Lvl2_c_094_2
  - Lvl2_c_095_1
  - Lvl2_c_096_5

DIENSTAG, 1.9.2026

- [x] overnight culture of: Ahmed Haj Kacem
  - Lvl2_c_094_2
  - Lvl2_c_095_1
  - Lvl2_c_096_5

MITTWOCH, 2.9.2026

- [x] glycerol stock of:
  - Lvl2_c_094_2
  - Lvl2_c_095_1
  - Lvl2_c_096_5

- [x] cPCR according to cPCR S. cerevisiae Taq Ahmed Haj Kacem
  - Lvl2_c_091
  - Lvl2_c_092
  - Lvl2_c_093
  - Lvl2_c_089 (Nanreaktor)

Master Mix Preparation Taq cPCR S. cerevisiae Level 2 modules: 89 & 91 - 93

	A	B	C
1		**25ul Reaction**	MM x 21.2
2			
3	2x Taq MM	12.5	265
4	10uM P fw	0.5	10.6
5	10um P rv	0.5	10.6
6	colony suspension	2	
7	H20	11.5	243.8

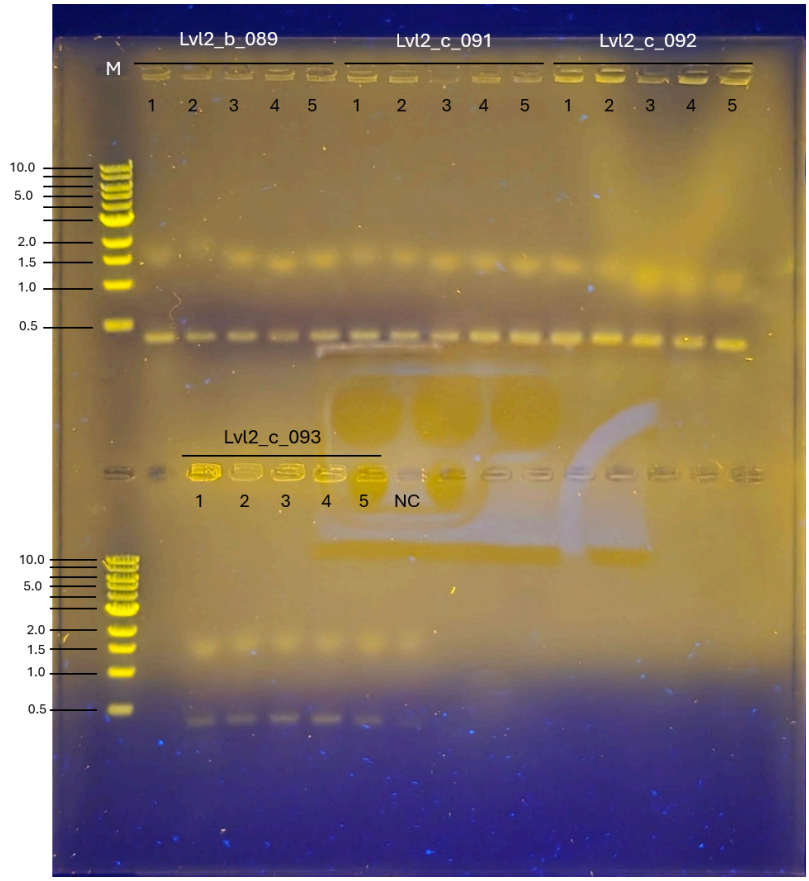
The size of the amplicon: 400 bp = 0.4 kb
elongation: 1 minute/kb
Annealing temperature 53 degree

cPCR S. cerevisiae cycle

	A	B	C
1	STEP	TEMP	TIME
2	Initial Denaturation	95°C	30 seconds
3	25-33 Cycles	95°C 45-68°C* 68°C	15-30 seconds 15-60 seconds 24 seconds
4	Final Extension	68°C	5 minutes
5	Hold	4-10°C	

- [x] gel electrophoresis of: Alex John Thottumkal
  - Lvl2_c_091
  - Lvl2_c_092
  - Lvl2_c_093
  - Lvl2_c_089 (Nanreaktor)

S. cerevisiae Taq cPCR: Lvl2_b_089, Lvl2_c_091-093



all are positive: any colony could be picked

DONNERSTAG, 3.9.2026

- [x] Miniprep of: Ziyue Zhou
  - Lvl2_c_094_2
  - Lvl2_c_095_1
  - Lvl2_c_096_5

miniprep 094-096

	A	B	C	D
1		c(ng/ul)	A260/230	A260/280
2	094_2	529	1.89	2.28
3	095_1	371	1.85	2.10
4	096_5	350	1.87	2.22

- [x] set up yeast preculture for trafo S. cerevisiae Transformation of: Ziyue Zhou
  - Lvl2_c_094_2
  - Lvl2_c_095_1
  - Lvl2_c_096_5

FREITAG, 4.9.2026

- [x] yeast trafo of S. cerevisiae Transformation Ziyue Zhou :
  - Lvl2_c_094_2
  - Lvl2_c_095_1
  - Lvl2_c_096_5
  - heat shock for 20min

SAMSTAG, 5.9.2026

- [x] set up precultures of: Ahmed Haj Kacem
  - Lvl2_c_091
  - Lvl2_c_092
  - Lvl2_c_093
  - Lvl2_c_089 (Nanreaktor)

  - the preculture was set up at ca. 9:30

MONTAG, 7.9.2026

- [x] create a masterplate of the picked colonies from the yeast transformed: Ahmed Haj Kacem
  - Lvl2_c_094
  - Lvl2_c_095
  - Lvl2_c_096
  - five colonies per construct were picked
- [x] induction of the precultures of: Purification of Encapsulin (S.cerevisiae) Ahmed Haj Kacem
  - Lvl2_c_091
  - Lvl2_c_092
  - Lvl2_c_093
  - Lvl2_c_089 (Nanreaktor)

The precultures were induced at 16:02. Let grow in induction medium until Tuesday 08/09/2026 14:02.

DIENSTAG, 8.9.2026

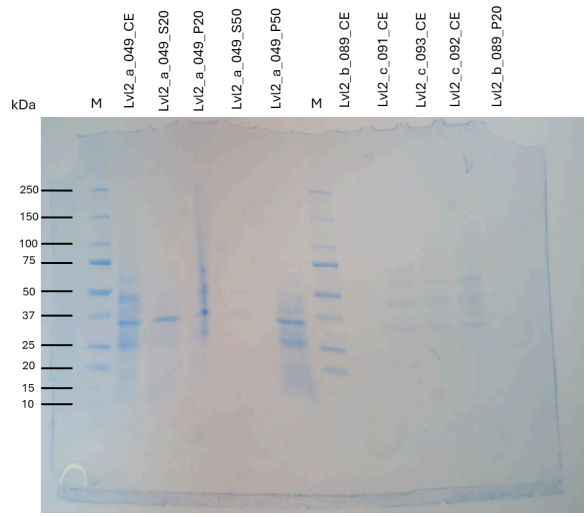
- [x] Cell harvest Ahmed Haj Kacem
- [x] precipitation, and Alex John Thottumkal
  - centrifugation following rocking after second precipation was performed without cooling
  - samples were frozen after last precipiation step to store for SDS-Page on the next day
  - (Learning for next time: best point in time to stop is right after harvest and before precipitation)

MITTWOCH, 9.9.2026

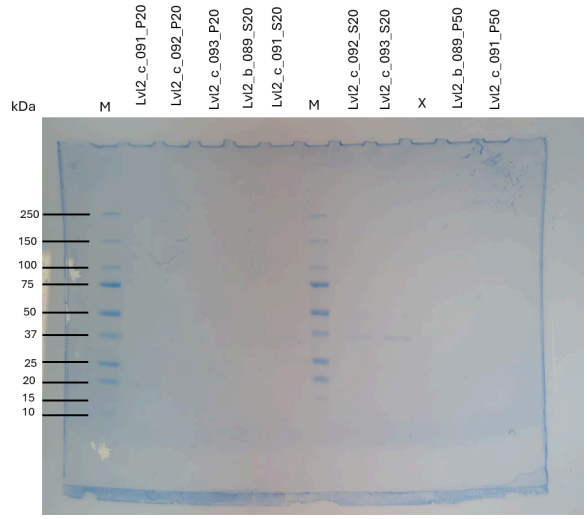
- [x] SDS-PAGE of: Ahmed Haj Kacem
  - Lvl2_c_091
  - Lvl2_c_092
  - Lvl2_c_093
  - Lvl2_c_089 (Nanreaktor)

  Results:

SDS-PAGE: Lvl2_a_049 (CE-P50) / Lvl2_b_089 (CE-P20) / Lvl2_c_091-093 (CE)



SDS-PAGE: P20: Lvl2_c_091-093, S20: Lvl2_b_089 & Lvl2_c_091-093, P50: Lvl2_b_089 & Lvl2_c_0



grad celsius